

High-plex spatial multiomic technology at the single-cell level in mouse FFPE brain tissue

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Abstract

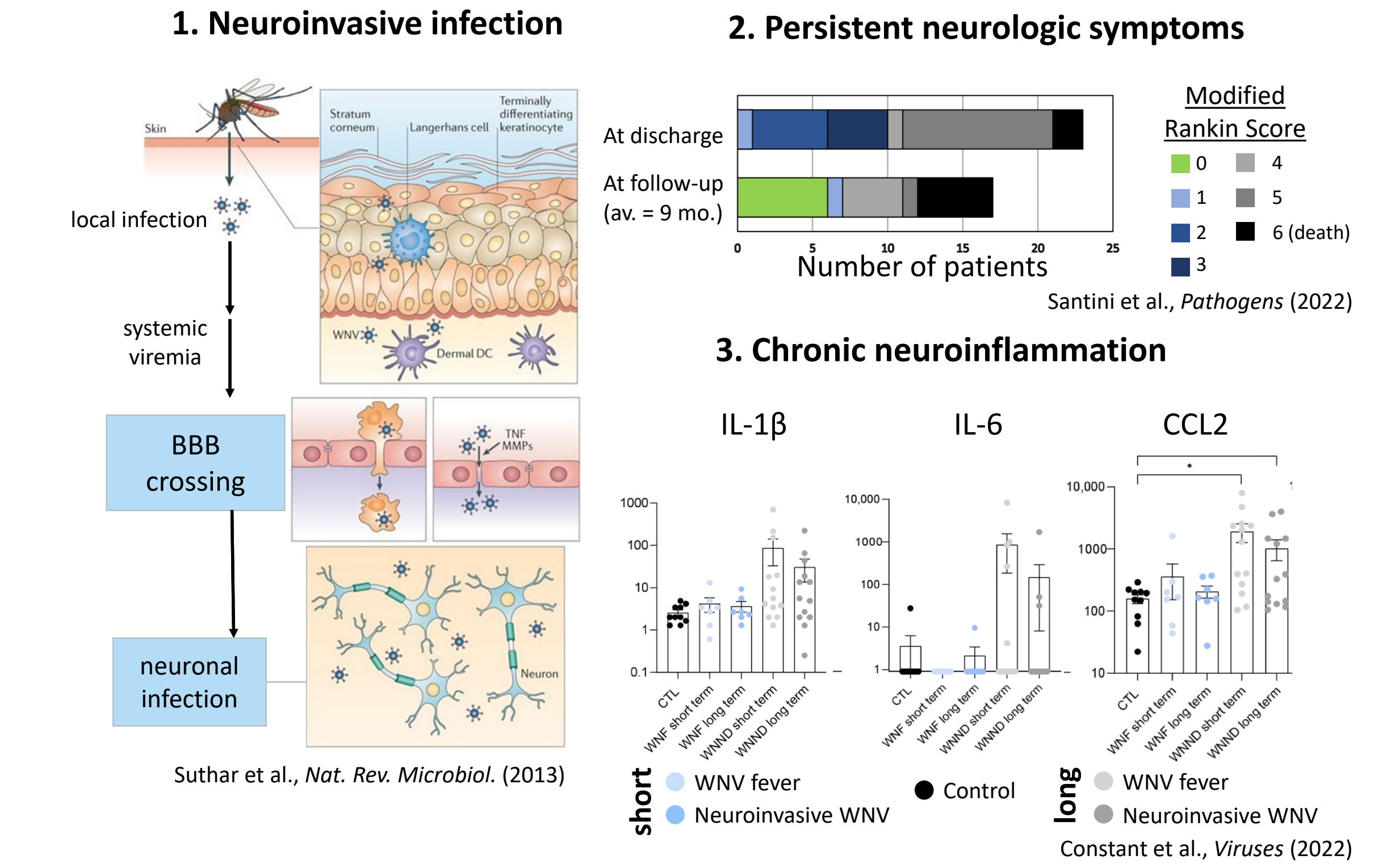
Spatially resolved, single-cell transcriptomics and proteomics in mouse neural models reveal neurobiological mechanisms relevant to human disease. However, spatial-omic protocols are analyte-specific and fail to capture multiomic information within the same single cell. We develop a multiomic workflow on a Spatial Molecular Imager (SMI), a single-cell spatial biology platform that leverages cyclic in situ hybridization chemistry to enable high-plex detection of proteins and RNAs at subcellular resolution. We measured 68 proteins using the Mouse Neuroscience protein panel (Neural Cell Typing and Alzheimer's Pathology) and 1,000 RNA targets (Mouse Neuroscience RNA panel) on the same 5 µm thick FFPE section of the mouse brain.

Our multiomic workflow demonstrates significant benefits to cell segmentation using cell-type specific markers (GFAP, Iba1, NeuN, in addition to a pan soma marker S6 and nuclear stain DAPI) in neural tissue. As a result, the number of transcripts captured within a single cell increased, most notably transcripts distal to cell bodies. This improved the quality of cell typing within the brain, including the detection of rare cell types and states.

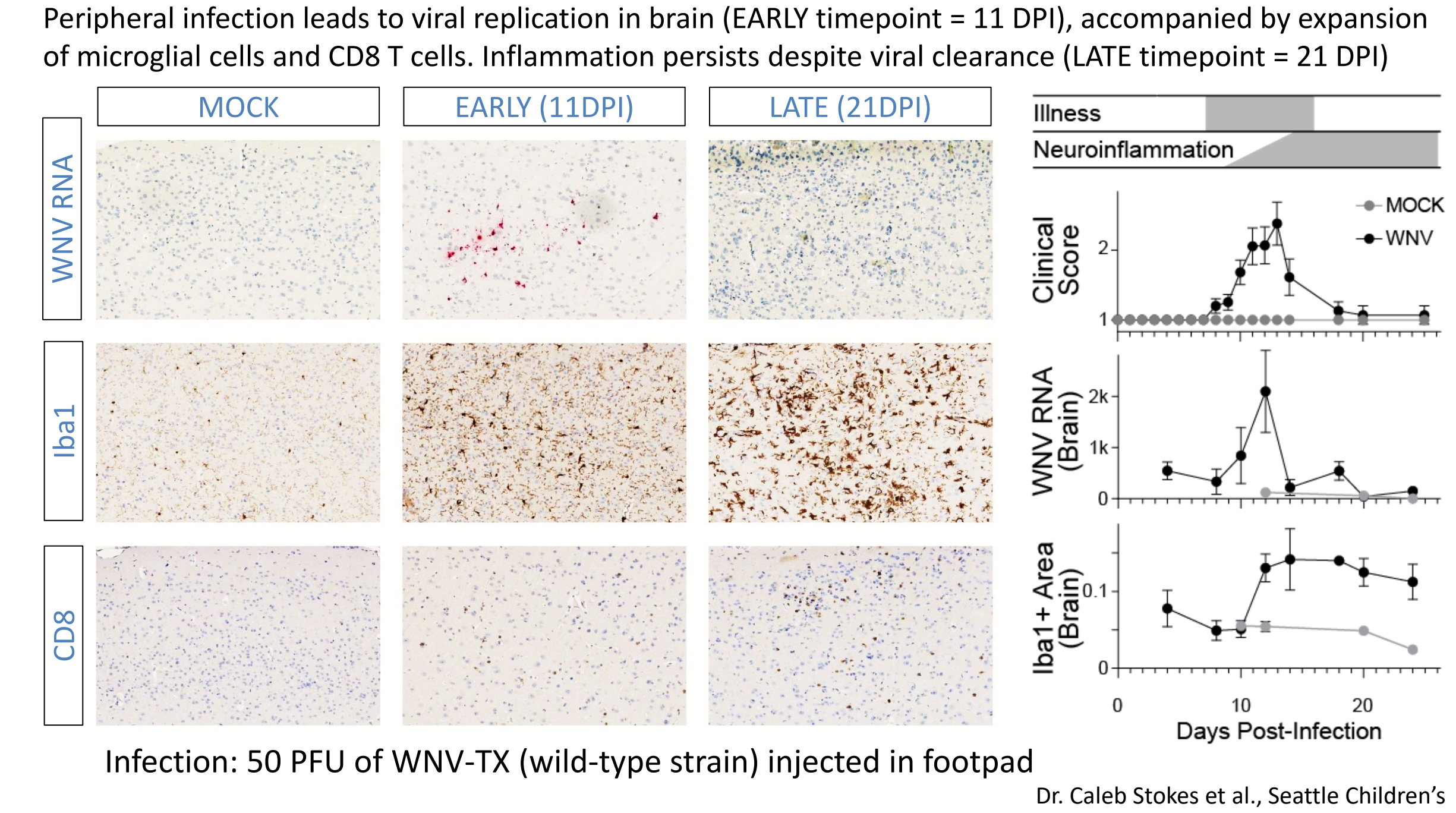
To validate the multiomic workflow, we profiled the uninfected mouse brain and the brain with West Nile Virus (WNV)-infected encephalitis. As expected from WNV encephalitis, we observed persistent neuroinflammation that includes the recruitment and activation of CD8+ T cells and microglia. We captured the major cell types that comprise inflammatory nodules in post-WNV mouse brain (neurons, microglia, and astrocytes) and identified the signaling pathways that underlie persistent microglial-driven neuroinflammation after WNV encephalitis, which illuminates key aspects of neurodegeneration, neurodevelopment, cell state and signaling, including numerous ligands and receptors involved in neuron-glia communication. Our analysis identified pathways related to inflammation and cellular damage in neurons, astrocytes, microglia and T cells.

Taken together, we used the CosMx™ SMI platform to show, for the first time, that large numbers of mouse neuronal cells can be profiled with both protein and RNA at single-cell resolution in a spatial context. This integrated system maximizes the information content per single cell to enable a mechanistic understanding of infectious disease pathology and inflammatory response in the brain.

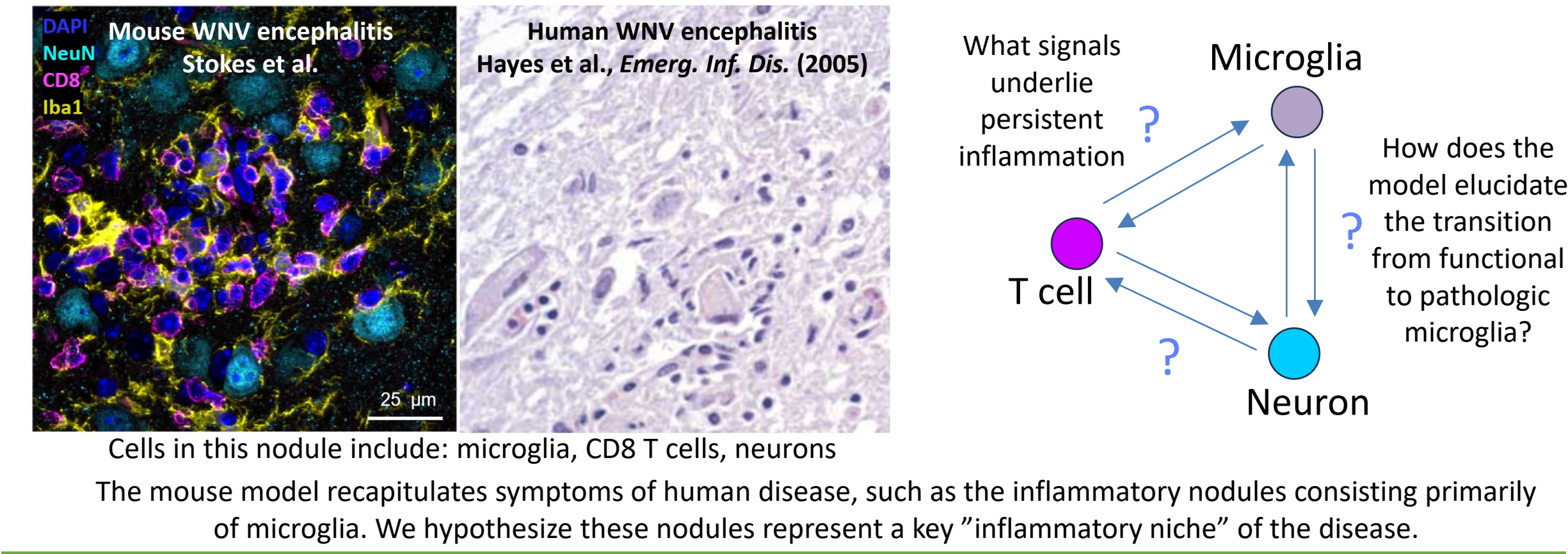
Neuroinflammation in West Nile Virus (WNV) Encephalitis



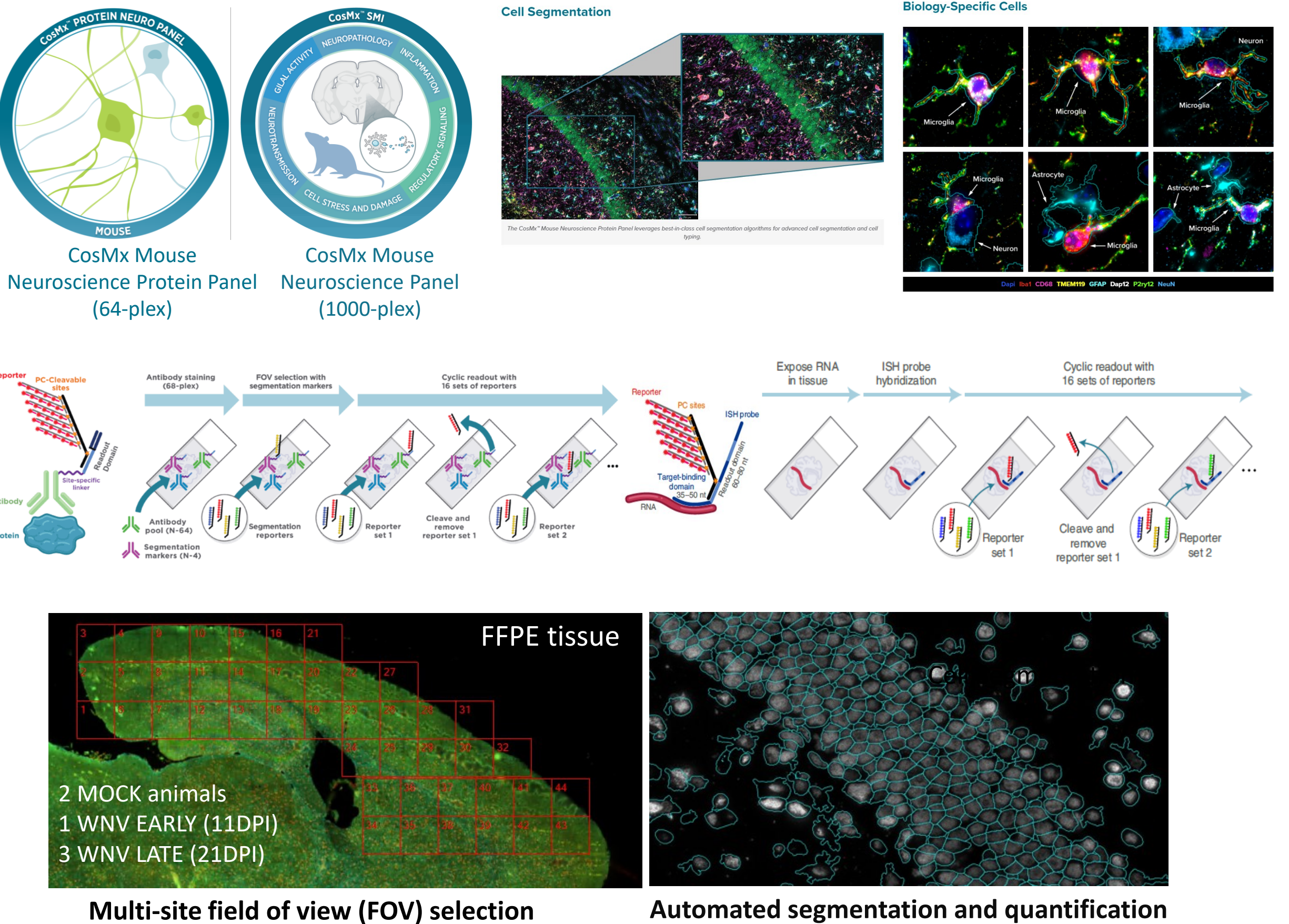
A Mouse Model of WNV-triggered Neuroinflammation



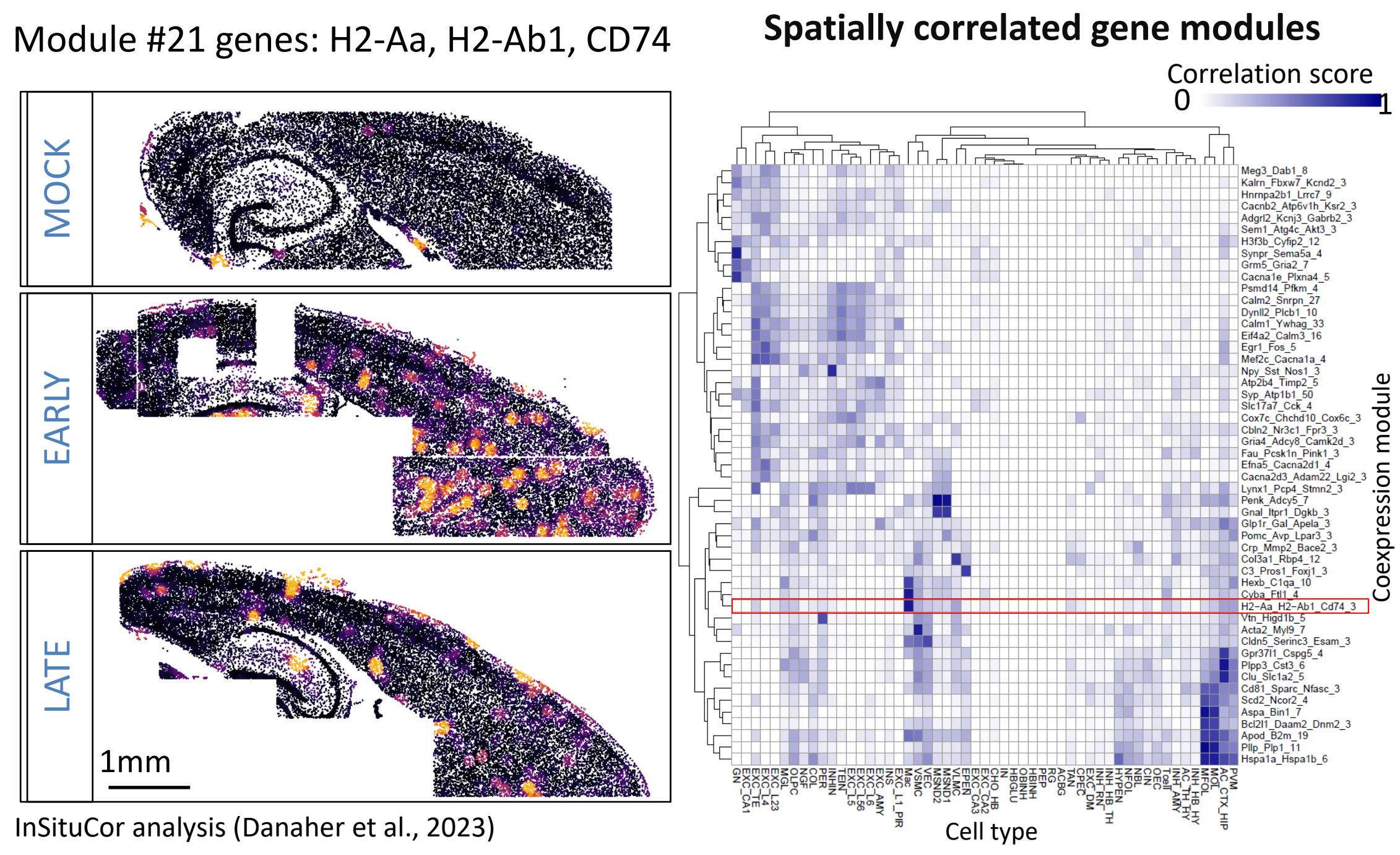
Nodular inflammation in WNV encephalitis



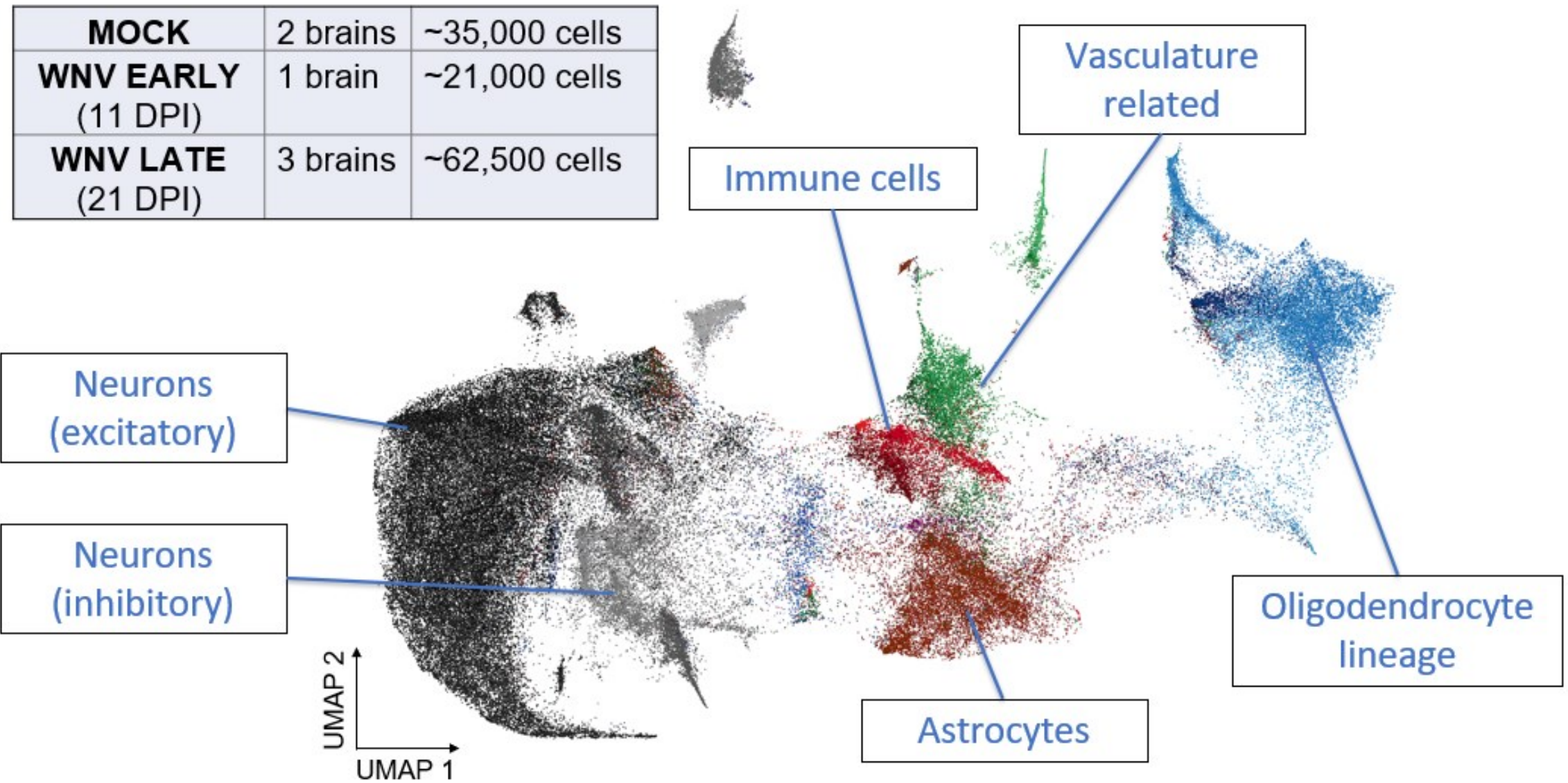
Approach: Multiomic Spatial Profiling of Inflammation in collaboration with Dr. Caleb Stokes at Seattle Children's



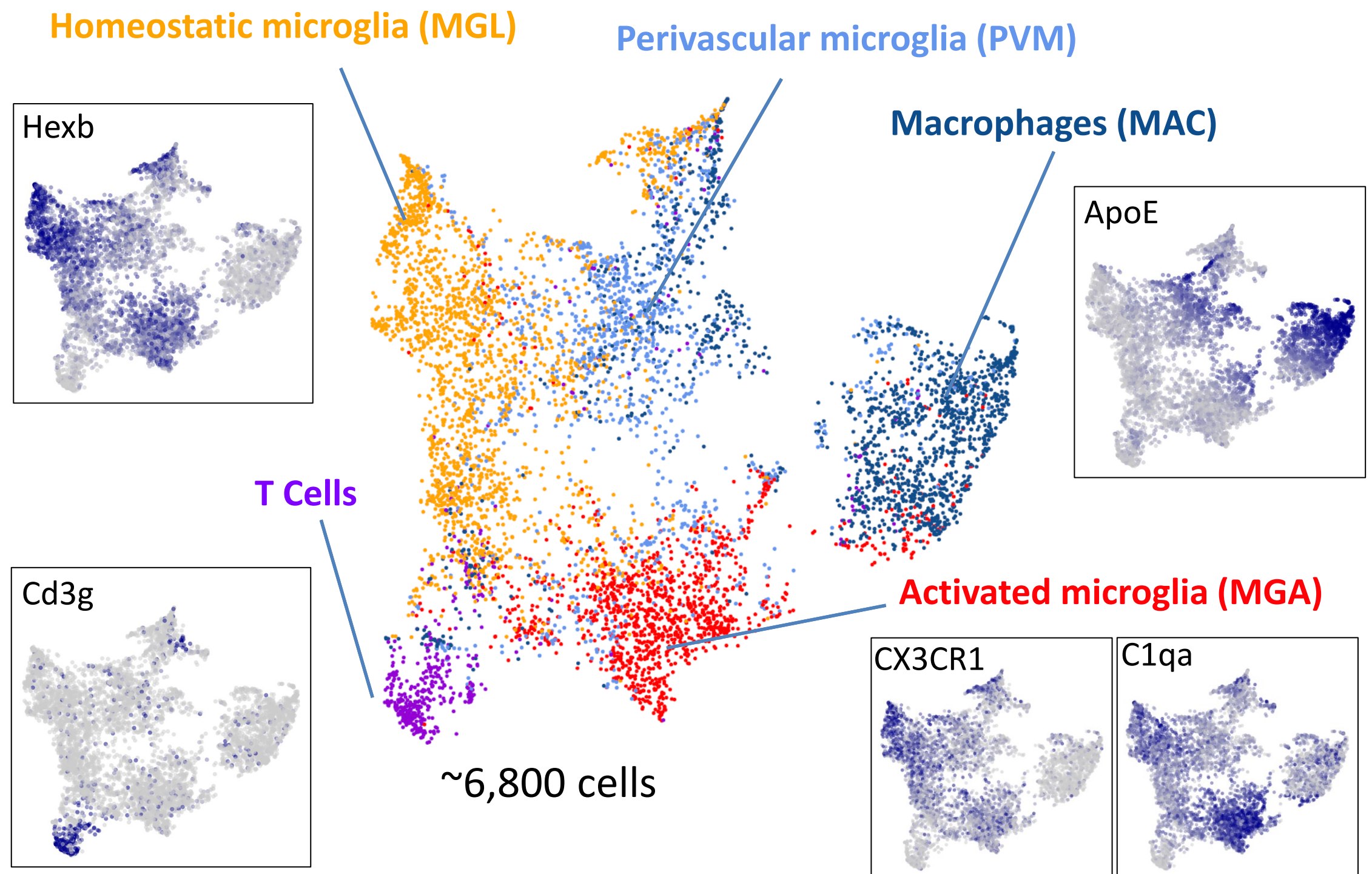
Inflammatory Transcripts are Spatially Circumscribed



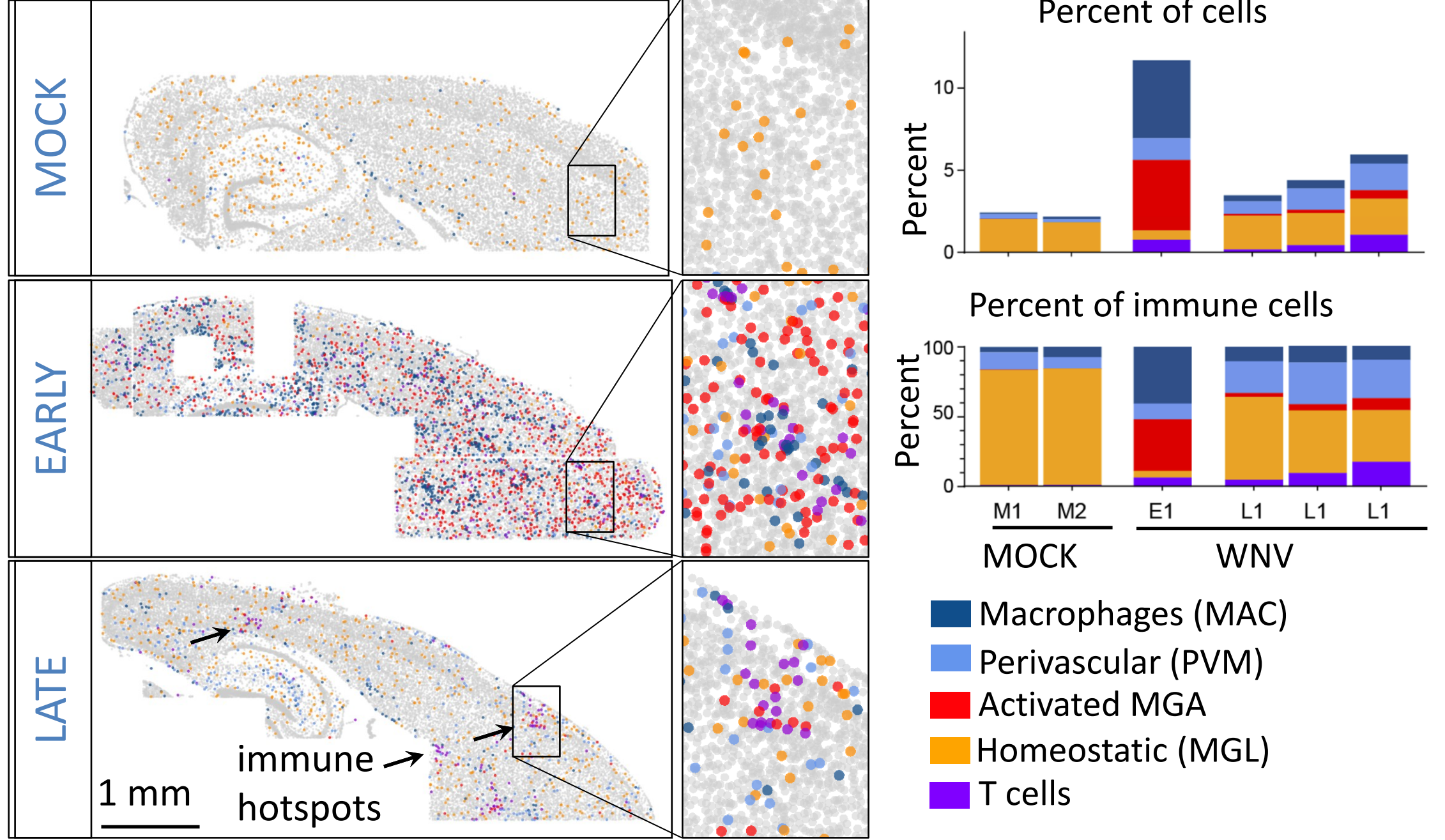
Transcription-based Clustering Identifies Major Cell Types



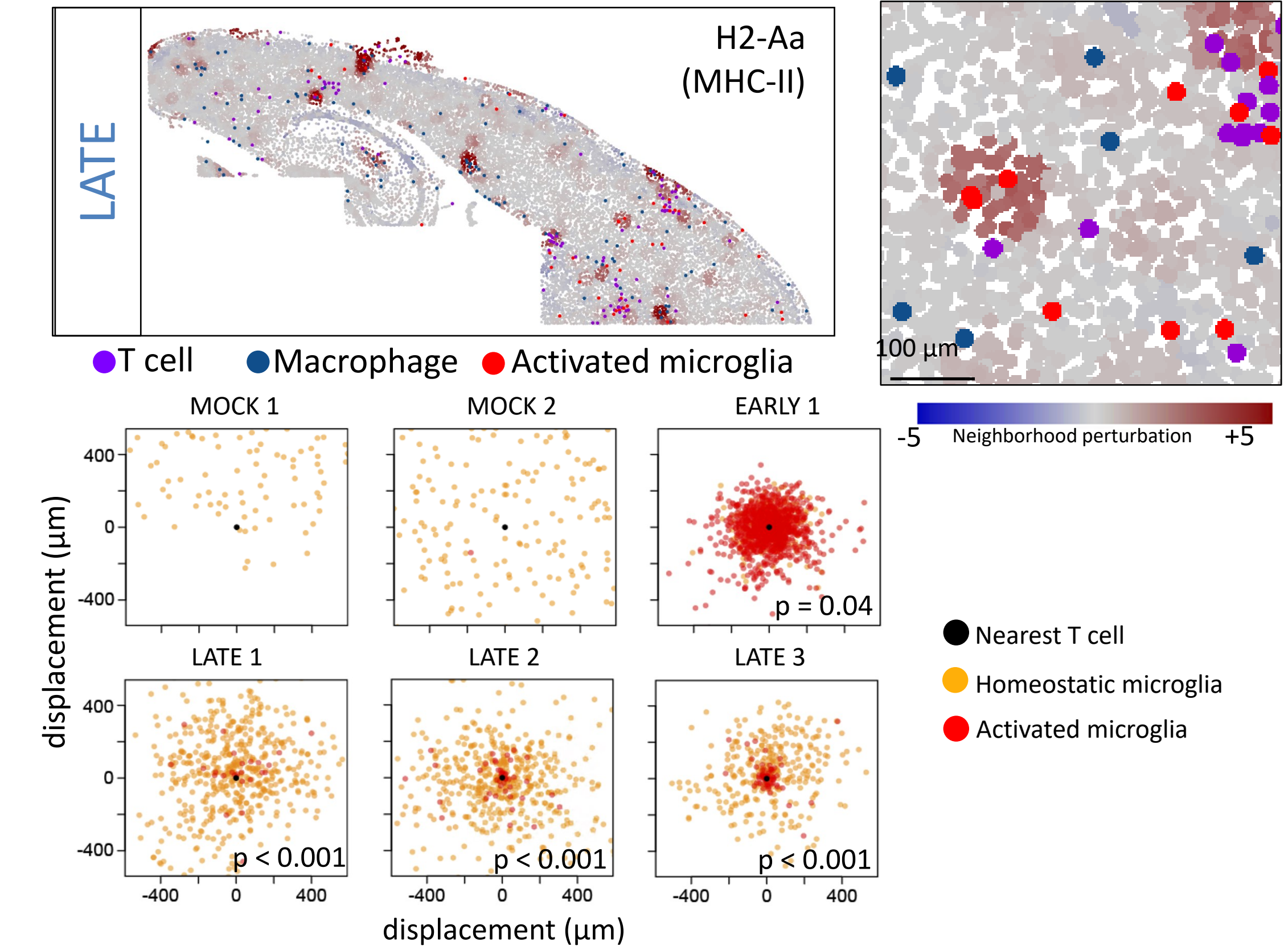
Diverse Immune Cell Populations in WNV Encephalitis



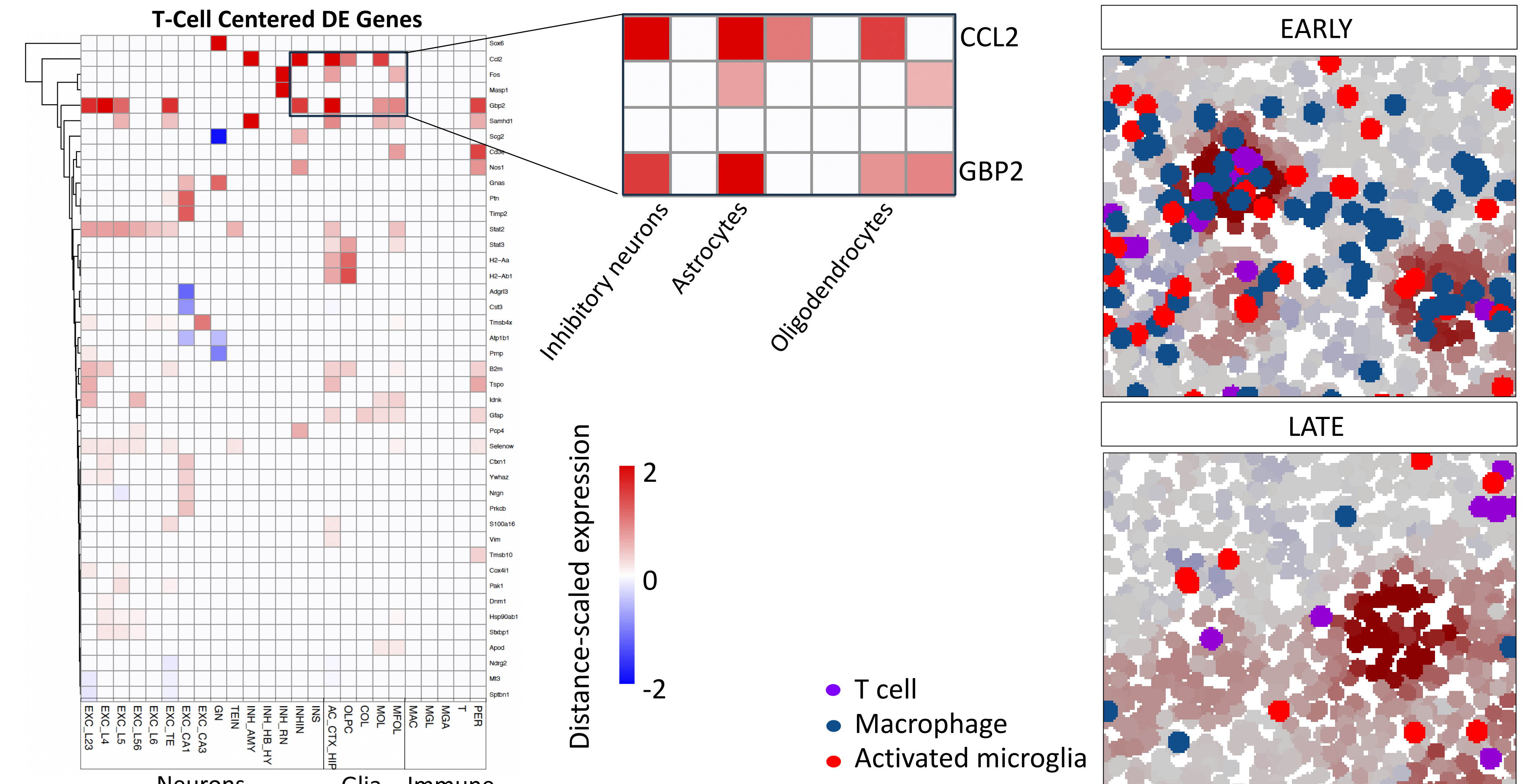
Spatial Population Dynamics of Immune Cells; Microglia Shift From Activated to Homeostatic in Later Stages



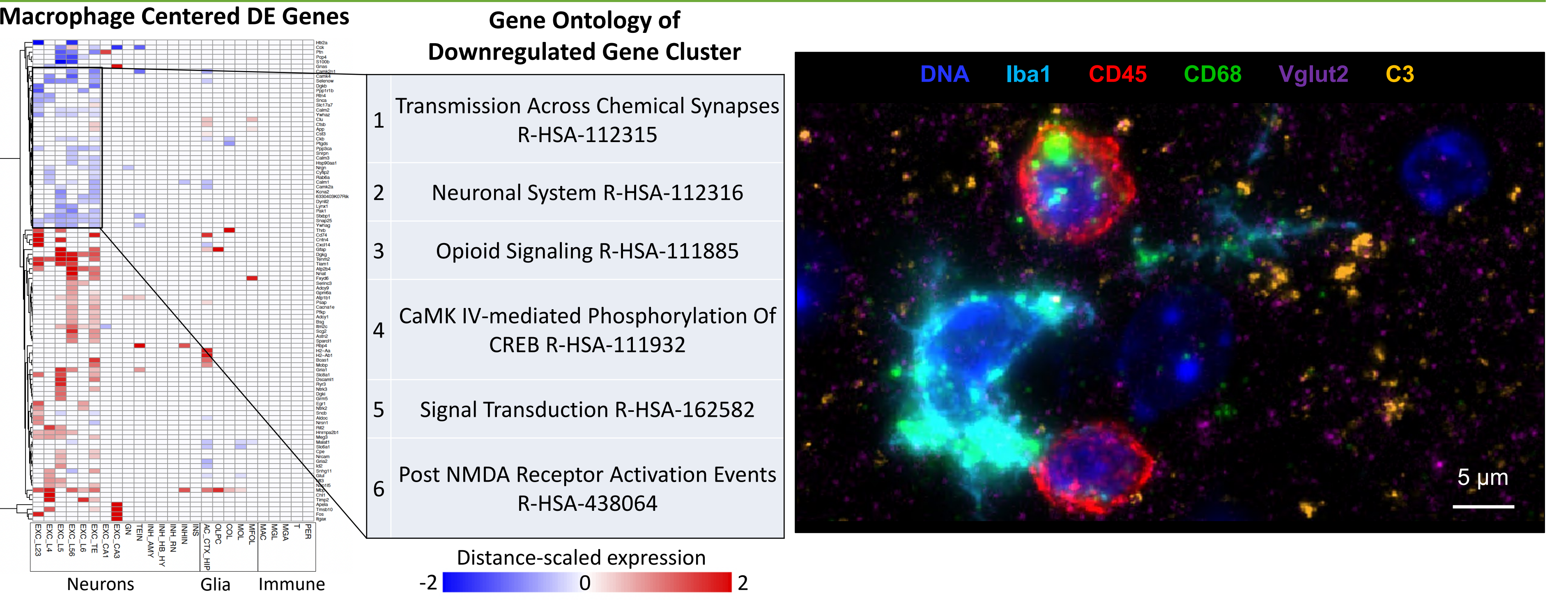
Activated Microglia Are Spatially Linked With T-Cells



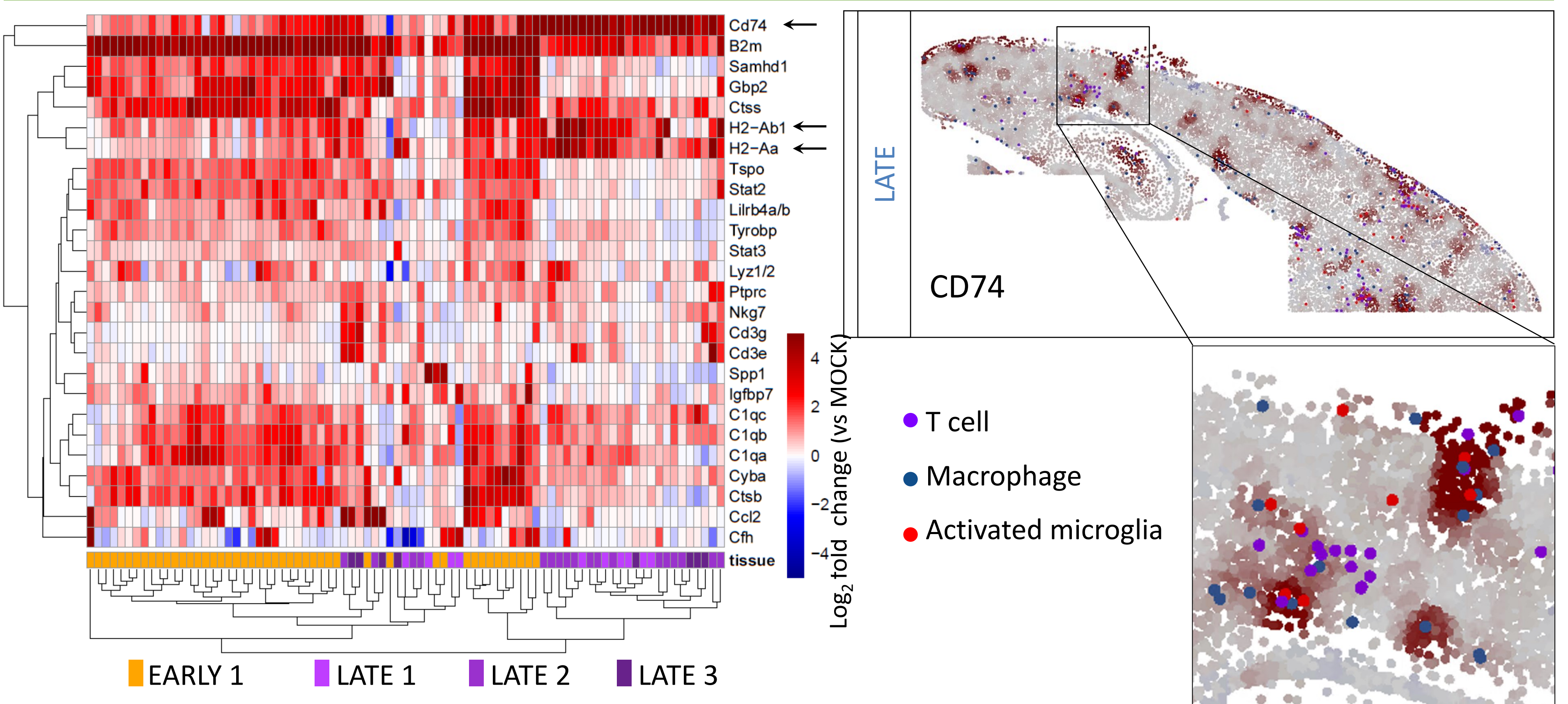
Co-expression of CCL2 and GBP2 in Inhibitory Neurons, Astrocytes, and Oligodendrocytes Drives T-Cell Recruitment in WNV Encephalitis



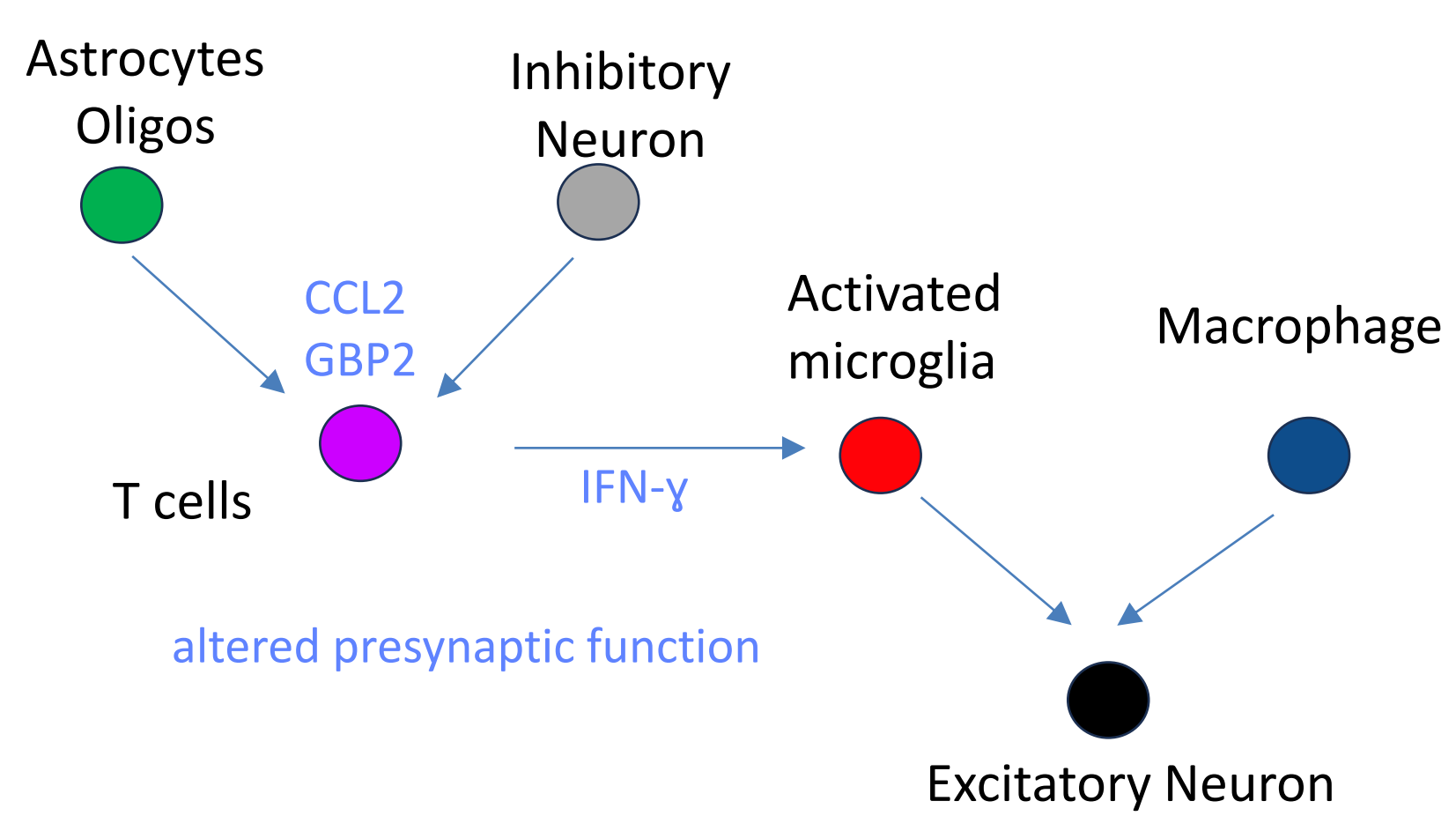
Downstream Consequences of Microglial Activity Include Downregulation of Genes Involved in Synaptic Transmission and Signaling Pathways



Gene Expression Changes Within Immune Hotspots



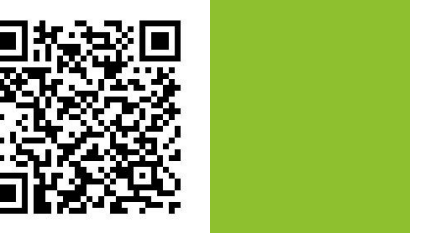
Drivers of Nodular Inflammation in WNV Encephalitis



Conclusions

- By utilizing a spatial multiomics approach, we:
- Profile 64 proteins and 1000 genes in a mouse model of WNV-triggered Neuroinflammation
 - Identify key cell types involved in the inflammatory response, including diverse immune cell populations
 - Reveal how spatial dynamics of immune cell populations change over time
 - Demonstrate a spatial relationship between activated microglia and T-cells
 - Reveal key gene expression signatures in neurons, astrocytes, and oligodendrocytes involved in the recruitment of T-cells
 - Implicate microglial activation in the downregulation of essential neuronal functions such as synaptic transmission and signaling pathways
 - Characterized changes in gene expression across the timeline of neuroinflammation, identifying a marked increase in CD74 at late stages

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